

I N S T R U C T I O N S

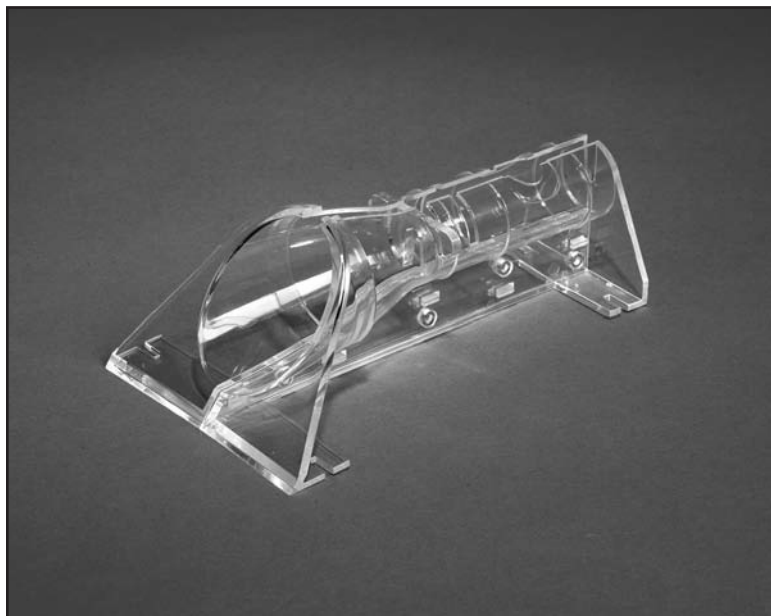


Mouse Restrainer with Dorsal Access

Catalog No. F46401-0000

Instructions for Use:

This restrainer is designed to hold mice with body weights between 18 and 35 grams, and can be used for tail bleeds, tail tattooing, and for dorsal subcutaneous injections. Inspect restrainer for any signs of damage before using. Place the restrainer on a clean flat surface. If needed, the restrainer can be held in place with fasteners applied through the four slots located on the bottom of each corner. Do not leave the mouse unattended.



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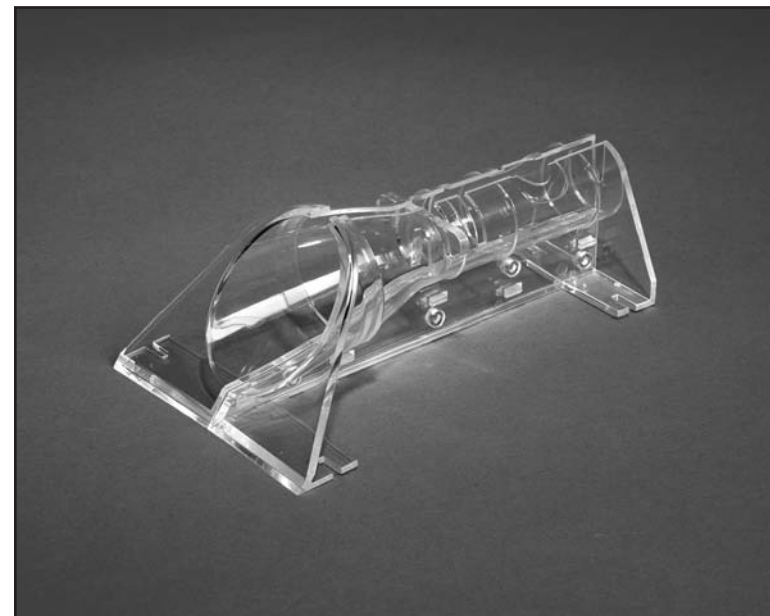


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For Dorsal Subcutaneous Injections:

1. Grasp the mouse by the base of the tail and gently draw the mouse tail first through the funnel, or lead head first into the wide entry funnel and to the back of the tube.
2. Place the included gate into one of the two available slots to secure the mouse inside the tube.
3. Use your forefinger and thumb to grasp the dorsal skin of the mouse through the beveled, circular recess on the top of the restrainer. This allows subcutaneous injections to be performed quickly, without risk of being bitten by mouse, and with reduced likelihood of sticking one's own fingers with needle.
4. To remove the mouse, grasp by the base of the tail then lift the gate.
Ease the animal out of the restrainer.

For Tail Bleeds and/or Tail Tattooing:

5. Insert the mouse tail first as described above (Step 1). Continue to draw the mouse to the end of the tube, allowing the tail to protrude through the opening at the end of the tube.
6. Place the included gate into one of the two available slots to secure the mouse inside the tube.
7. Tail bleeds and/or tail tattooing can now be performed without risk of being bitten and without risk of collateral damage to the mouse.
8. To remove the mouse, grasp by the base of the tail then lift the gate.
Ease the animal out of the restrainer.

Cleaning and Disinfecting the Restrainer:

All parts of the restrainer are made from TPX® polymethylpentene, a clear, autoclave-safe, and chemical-resistant plastic material.

1. Remove and wipe any bodily excretions from the mouse that may be present in or on the restrainer. Dispose in a manner consistent with all local, state and federal regulations.
2. The restrainer may be cleaned using most laboratory glassware/plasticware cleaners. We recommend using Aquet® (Scienceware® item nos. F17094-0020 or F17094-0030), a non-ionic, phosphate-free, biodegradable surfactant that will not etch or mar the restrainer.
3. Once cleaned, the restrainer may be disinfected by dipping in alcohol or similar commonly used laboratory disinfectants.

Autoclaving Instructions:

The restrainer may be steam autoclaved as needed. It is recommended to wash product before autoclaving to prevent baking contaminants into the surface of the plastic during autoclave process. Recommended autoclave cycle is 121°C at 15 psig (1 bar) for 20 minutes.



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6. Place the included gate into one of the two available slots to secure the mouse inside the tube.
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